



Glutamate hypothesis in schizophrenia

Yota Uno, MD, PhD^{1,2,3*} and Joseph T. Coyle, MD^{1,2}

Schizophrenia is a chronic and severe psychiatric disorder that has profound impact on an individual's life and on society. Thus, developing more effective therapeutic interventions is essential. Over the past quarter-century, an abundance of evidence from pharmacologic challenges, post-mortem studies, brain imaging, and genetic studies supports the role of glutamatergic dysregulation in the pathophysiology of schizophrenia, and the results of recent randomized clinical trials based on this evidence have yielded promising results. In this article, we review the evidence that alterations in glutamatergic neurotransmission, especially focusing on the *N*-methyl-D-aspartate receptor (NMDAR)

function, may be a critical causative feature of schizophrenia, how this contributes to pathologic circuit function in the brain, and how these insights are revealing whole new avenues for treatment development that could reduce treatment-resistant symptoms, which account for persistent disability.

Keywords: *D*-serine, glutamate, *N*-methyl-D-aspartate receptor, schizophrenia, serine racemase.

<http://onlinelibrary.wiley.com/doi/10.1111/pcn.12823/full>

Schizophrenia is a chronic and severe psychiatric disorder that has a profound impact on both an individual's life and on society. It is characterized clinically by positive symptoms (i.e., hallucinations, delusions, disorganized thinking, and grossly disorganized or abnormal motor behavior), negative symptoms (i.e., diminished emotional expression, avolition, apathy, anhedonia, and social withdrawal), and cognitive deficits (i.e., deficits in attention, working memory, and executive function). The cognitive deficits are known to appear before clinical diagnosis¹ and are strongly linked to functional outcomes, such as academic and occupational function and independent living.

The prevalence of schizophrenia has been estimated to be approximately 1% in the general population, and it exhibits high heritability. According to a recent twin study with a large, adequately powered sample and improved statistical methods,² the heritability was calculated to be 79%, consistent with prior results. However, the pattern of inheritance is non-Mendelian, supporting the role of complex genetics. Thus, genetic risk factors strongly contribute to the cause of schizophrenia, but at the same time environmental factors also play a significant role. Schizophrenia is considered to be a neurodevelopmental disorder because environmental insults, such as infection, are concentrated in the perinatal period.³

In this article, we will review the mounting evidence that dysfunction of the *N*-methyl-D-aspartate receptor (NMDAR) may be a critical causative feature of schizophrenia, how this contributes to pathologic circuit function in the corticolimbic system, and how these insights are revealing whole new avenues for treatment development that could ameliorate the current treatment-resistant negative and cognitive symptoms, which account for persistent disability.

The history of pathological hypotheses in schizophrenia

Hypotheses before the discovery of antipsychotic drugs

Existence of people with psychiatric symptoms, such as hallucinations and delusions (phrenitis), was already known in the era of Hippocrates (460–375 BC), the so-called father of medicine. His

contemporaries believed that these disorders were due to abnormal conditions of the body caused by physical factors. They tried to treat these disorders in several ways believed to have potential therapeutic effects, including eating the root of *Rauvolfia serpentina*, which contains reserpine that inhibits the storage and use of biogenic amines.

In 1899, Emil Kraepelin, a German psychiatrist, integrated several psychiatric conditions, such as hebephrenia, catatonia, and dementia paranoides, which are now known to be subtypes of schizophrenia, and developed the concept of dementia praecox.⁴ Since his assumptions were that those conditions led to irreversible loss of cognitive functions and were resistant to treatment, he classified them as a dementia.

The term 'schizophrenia,' which is used currently, was coined in 1908 by Eugen Bleuler, a Swiss psychiatrist.⁵ He revised the idea of dementia praecox and renamed it 'schizophrenia.' Notably, he identified a disconnect between affect and thought processes (splitting, e.g., 'schiz') as the most fundamental feature of schizophrenia and ambivalence and autism as basic symptoms, whereas positive symptoms, such as hallucinations and delusions, were considered as accessory symptoms.^{6,7} He also speculated that the core symptoms may respond to different treatments than the ancillary symptoms. This hypothesis proved prescient when antipsychotics were found to be effective against positive symptoms but ineffective against negative and cognitive symptoms.⁸ Later, Kurt Schneider, a German psychiatrist, proposed a list of highly disorder-specific forms of delusions and hallucinations based on a disturbance of self-identity, which he termed 'first-rank symptoms,' whereas he deemed negative and cognitive symptoms as secondary symptoms.^{4,9} However, the specificity of first-rank symptoms for schizophrenia has been questioned.¹⁰

Dopamine hypothesis

The first antipsychotic, chlorpromazine, which was originally developed as an antihistaminergic drug for use in a range of conditions, including nausea and allergies, and reserpine, a *Rauvolfia* alkaloid, were demonstrated to be effective in treating the psychosis of

¹ Department of Psychiatry, Harvard Medical School, Boston, USA

² Laboratory for Psychiatric and Molecular Neuroscience, McLean Hospital, Belmont, USA

³ Department of Psychology, University of Bath, Bath, UK

* Correspondence: Email: yota_u@ypdc.net

schizophrenia^{11–13} in the early 1950s. Shortly afterward, more potent phenothiazines and butyrophenones drugs were developed and proved to be highly efficacious in alleviating symptoms of schizophrenia.^{14–16} However, they were prone to inducing Parkinsonian-like extra-pyramidal side-effects.¹⁵ Moreover, reserpine was found to cause depletion of monoamine, such as dopamine, in synaptic vesicles via inhibiting vesicular monoamine transporter. Alpha-methyl-para-tyrosine, which was known as a specific inhibitor of tyrosine hydroxylase, the initial enzyme in the synthesis pathway for catecholamines, was also shown to reduce symptoms of schizophrenia.¹⁷ Carlsson synthesized these observations and proposed that antipsychotics exerted their therapeutic effects by blocking dopamine receptors, for which he received the Nobel Prize.¹⁸

‘Stimulants,’ such as amphetamine, which inhibits the presynaptic vesicular monoamine transporter 2 and the dopamine transporter, increase dopamine concentrations in the synaptic cleft. Consistent with the antipsychotic effects of depleting dopamine, high doses of stimulants induce a psychosis that clinically resembles the acute phase of paranoid schizophrenia.¹⁹ Animal data indicated that both phenothiazines and butyrophenones reduced dopamine-mediated behaviors induced by amphetamine or apomorphine.^{20–22} Synthesizing the opposing effects of stimulants and antipsychotics on dopaminergic neurotransmission, Snyder proposed the ‘dopamine hypothesis’ that schizophrenia was due to excessive stimulation of dopamine receptors.²³

Glutamate hypothesis

The dopamine hypothesis can account for certain aspects of the psychopathology of schizophrenia, especially positive symptoms.²⁴ However, with the possible exception of clozapine, antipsychotics have negligible effects on negative and cognitive symptoms, the most robust predictors of disability in schizophrenia.²⁵ Furthermore, cortical atrophy correlates with negative and cognitive symptoms in chronic schizophrenia but not with the severity of psychosis.^{26,27} Thus, the core features of schizophrenia, which are primarily responsible for persistent disability, are linked to pervasive cortical pathology and are unlikely the consequence of simply dopamine dysfunction.²⁸

Ketamine and phencyclidine (PCP), which were developed as dissociative anesthetics in 1970s, have been known to induce schizophrenic-like symptoms, including not only psychotic symptoms and thought disorders but also negative and cognitive symptoms in healthy humans.^{29,30} Moreover, psychosis induced by ketamine or PCP is clinically difficult to distinguish from the primary psychosis of schizophrenia.^{31,32}

The existence of binding sites for ketamine and PCP were described in 1979 followed by reports in the early 1980s that they are noncompetitive antagonists of the NMDA subtype of glutamate receptor.³³ Based on clinical observations of patients intoxicated with PCP and ketamine and careful laboratory studies of the effects of infusion of subanesthetic doses of ketamine in normal volunteers. It was proposed that schizophrenia results from hypofunction of NMDAR.^{30,32,34}

What is the NMDA Receptor?

Glutamate is recognized as the most abundant excitatory amino acid neurotransmitter in the brain. It activates G protein-coupled metabotropic (mGlu) receptors and ionotropic receptors. The ionotropic receptors are divided into three subtypes based on their sensitivity to high-affinity selective ligands: NMDAR, α -amino-3-hydroxy-5-methylisoxazole-4-propionate receptors, and kainite receptors. NMDAR show relatively slow and incomplete desensitization compared to non-NMDA ionotropic glutamate receptors.

Structure and function (see Fig. 1)

NMDAR comprise a heterotetrameric complex of two obligatory GluN1 subunits with either two GluN2 subunits or a combination of

GluN2 and GluN3 subunits. The GluN1 subunit is encoded by a single gene (*GRIN1*) but has eight different isoforms owing to alternative splicing. GluN2 subunits and GluN3 subunits also have four (GluN2A - D) and two variants (GluN3A - B), which are encoded by separate genes, *GRIN2A-D* and *GRIN3A-B*, respectively. Each GluN subunit has a typical modular architecture with two large clamshell-like extracellular domains (an amino-terminal domain involved in assembly and channel modulation, and a ligand-binding domain), a transmembrane domain, and a carboxy-terminal domain involved in receptor trafficking and signaling. The amino-terminal domain and carboxy-terminal domain regions are the most divergent and account for much of the functional diversity of NMDAR.³⁵ D-serine and glycine bind to GluN1 and GluN3 subunits, which is designated the glycine modulatory site (GMS), and glutamate binds to GluN2 subunits at the glutamate binding site.³⁶ This receptor is modulated by the GluN2A and GluN2B allosteric antagonists divalent zinc and the phenylethanolamines ifenprodil or Ro25-6981, respectively.³⁷ Moreover, there are binding sites for adjusting ion channel activation, such as the redox modulatory site where glutathione binds on GluN1 and 2A,³⁸ spermine binding site on GluN2B and Mg^{2+} , MK-801, and the ketamine and PCP binding site on the transmembrane domain.³⁵

Activation of the NMDAR uniquely requires three simultaneous events: (i) postsynaptic depolarization, typically by activation of α -amino-3-hydroxy-5-methylisoxazole-4-propionate receptors, to remove the magnesium block of the GluN1 cation channel; (ii) occupancy of the GMS on GluN1 by either glycine or D-serine; and (iii) binding of the neurotransmitter glutamate to its receptor on GluN2 to open the channel and permit calcium entry. Calcium induces a cascade of intracellular events that mediate local, acute functional synaptic plasticity and changes in gene expression that promote long-term neural structural plasticity.³⁹

Pathophysiological Role of Serine Racemase and D-Serine

Vertebrates were thought not to utilize D-amino acids, although bacteria were known to synthesize several D-amino acids. Serine racemase (SR) and D-serine were discovered in eukaryotic insects in 1966.⁴⁰ In 1992, the Nishikawa laboratory was the first to report the presence of free D-serine in the mammalian brain and overthrew the shibboleth that vertebrates do not synthesize D-amino acids.⁴¹ This discovery also resolved a quarter-of-a-century conundrum about the reason for the expression of D-amino acid oxidase (DAAO), an enzyme that de-aminates D-amino acid to imino acid in the brain.⁴²

The Snyder laboratory developed the evidence that functional activity of NMDAR required endogenous D-serine as a co-agonist by showing loss of NMDAR activity by perfusing acute hippocampal slices with purified DAAO, which degraded synaptic D-serine, but not glycine.⁴³ Furthermore, they demonstrated that there was a close correspondence in the expression of SR, NMDAR, especially glutamate, glycine, and PCP binding sites, and D-serine levels in the forebrain^{44,45} and an inverse association in DAAO expression and D-serine levels in the cerebellum and hindbrain where glycine was found in high concentrations.²⁸ Thus, they concluded that D-serine is the primary co-agonist at synaptic NMDAR in the forebrain.^{28,46,47} They succeeded in the purification and characterization of SR from the rat brain in 1999.⁴⁸ This enzyme is classified as a member of the family of pyridoxal-5'-phosphate-dependent enzymes and catalyzes the formation of D-serine from L-serine.

Cellular localization

D-serine is enriched in corticolimbic regions of the brain and localized to the same areas as NMDAR.⁴⁴ The cellular source of D-serine has been hotly disputed. In initial *in vitro* studies,⁴⁹ D-serine was believed to be enriched in astrocytes and mainly synthesized in astrocytes, and therefore, SR was considered an astrocytic enzyme.^{44,48} While there is still a widely held belief that D-serine and SR are

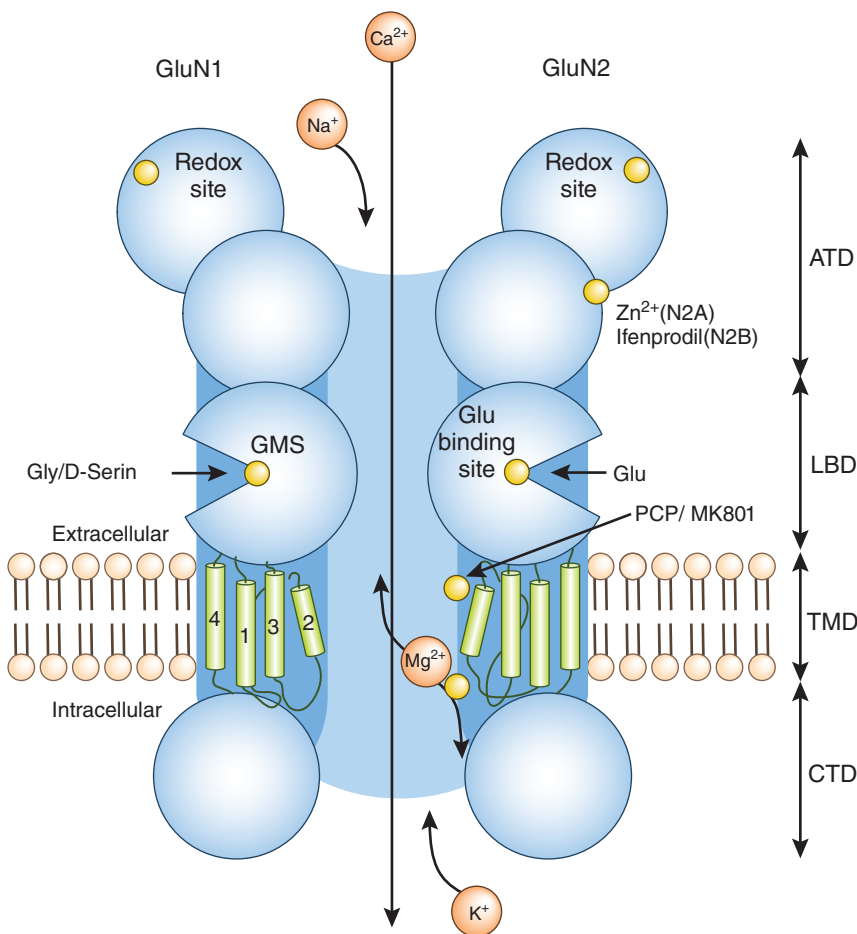


Fig.1 Structure of the *N*-methyl-D-aspartate receptor (NMDAR; GluN1/GluN2). The conventional NMDAR comprises a heterotetrameric complex of two GluN1 and two GluN2 subunits. ATD, amino-terminal domain; CTD, carboxy-terminal domain; GMS, glycine modulatory site; LBD, ligand-binding domain; PCP, phencyclidine; TMD, transmembrane domain.

components of a glial transmitter system,⁵⁰ immunohistochemical studies show that the majority of SR is expressed in neurons, rather than in astrocytes. For example, the Mori laboratory called into question the glial synthesis of D-serine a decade ago.⁵¹ In their study, they reported that SR immunoreactivity was predominantly localized to neurons: pyramidal neurons in the cerebral cortex and hippocampal CA1 region, medium-spiny neurons in the striatum and weakly gamma-aminobutyric acid (GABA)ergic Purkinje cells in the cerebellum by using novel SR knockout mice as a control for immunospecificity of their SR antibodies. Furthermore, using mice with conditional deletions of SR either in excitatory forebrain glutamatergic neurons or glial fibrillary acid protein-expressing astrocytes, Benneyworth *et al.* reported that the majority of SR was expressed in glutamatergic neurons and that, by contrast, less than 15% was in astrocytes in the hippocampus and none in the cerebral cortex.⁵² Moreover, in human post-mortem neocortex, SR was also found in both excitatory and inhibitory neurons, but not in astrocytes.⁵³

Recently, inflammatory A1 reactive astrocytes were found to express SR and synthesize and release D-serine. Reactive astrocytes typically proliferate after brain insults, such as traumatic brain injury (TBI). Hippocampal TBI was shown to cause proliferation of SR-expressing reactive astrocytes over 7 days while neuronal SR in injured neurons declined.⁵⁴ Notably, silencing the expression of SR in astrocytes by conditional *srr* inactivation only in astrocytes prevented the electrophysiologic and cognitive deficits after TBI. D-Serine released from reactive astrocytes would preferentially act at extrasynaptic neuronal NMDAR, which cause excitotoxic neuronal damage and death. Consistent with the TBI findings, primary cultures of astrocytes obtained from neonatal mouse brain express markers associated with reactive astrocytes and also express SR and release D-serine.^{55,56}

D-Serine homeostasis: The serine shuttle (see Fig. 2)

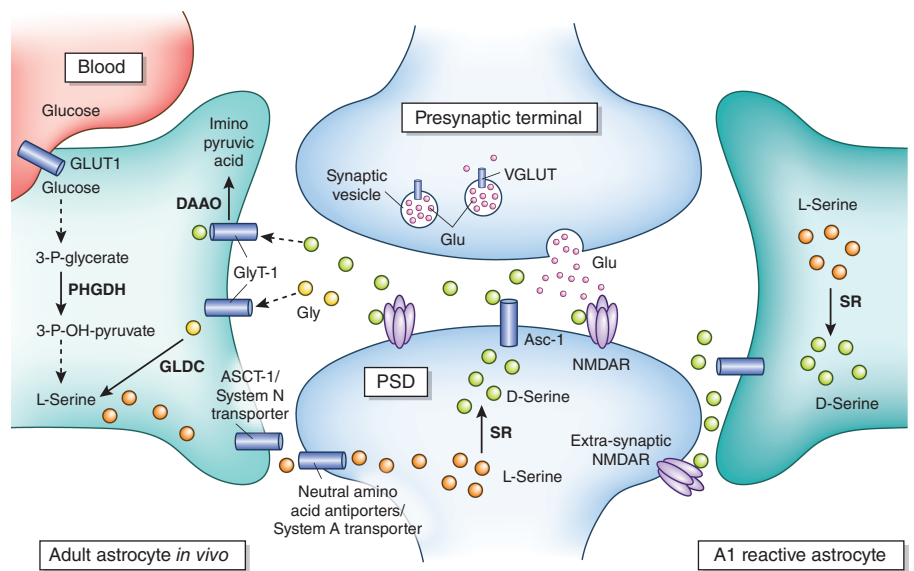
Recent studies indicate that resting adult astrocytes *in vivo* are the primary source of brain L-serine and express all the necessary enzymes to convert glucose into L-serine.^{57,58} SR and D-serine are co-localized with postsynaptic density protein 95 (PSD-95) at postsynaptic densities in glutamatergic synapses on both excitatory and inhibitory neurons, but not within the presynaptic vesicular glutamate transporter-1.⁴⁶ Deletion of Phgdh in mouse brain astrocytes dramatically reduces brain L- and D-serine by about 80%,⁵⁸ with D-serine deficits localized to neurons.⁵⁹ Thus, L-serine is considered to be synthesized from glucose via a 3-phosphoglycerate dehydrogenase-dependent pathway in adult astrocytes and then transported into postsynaptic neurons to be converted to D-serine by neuronal SR.⁴⁹ This process has been designated the 'serine shuttle,' which highlights the role of glia–neuron cross-talk for optimal NMDAR function.⁶⁰ As D-serine is released from postsynaptic spines at glutamatergic synapses, D-serine appears to function in an autocrine fashion and not as a presynaptically localized co-transmitter. This autocrine property of D-serine ensures that the postsynaptic NMDAR's GMS are occupied with D-serine to respond immediately to released glutamate.²⁸ Termination of D-serine signaling is attained by reuptake into astrocytes where it is catabolized by DAAO. On the other hand, under pathologic conditions, A1 reactive astrocytes express SR and release D-serine, which may contribute to their neurotoxic effects by acting at extrasynaptic NMDAR.^{28,56}

Evidence Supporting the NMDAR Hypothesis

Clinical findings

The most impressive clinical evidence that PCP psychosis resembles schizophrenia is the fact that PCP users have been mistaken by experienced psychiatrists for having schizophrenia before obtaining the

Fig.2 Schematic representation of serine shuttle. L-serine is synthesized from glucose, which diffuses from blood vessels and is transported into astrocytes by glucose transporter 1 (GLUT1). L-serine is then transported into postsynaptic neurons via neutral amino acid exchangers (such as ASCT1, neutral amino acid antiporters, or system A transporter) to be converted to D-serine by serine racemase (SR). This process is known as the 'serine shuttle.' SR and D-serine are concentrated at the postsynaptic densities in glutamatergic and SR-expressing GABAergic neurons. D-serine is subsequently released from neurons by Asc-1 or other transporters and binds to the glycine modulatory sites on synaptic N-methyl-D-aspartate receptor (NMDAR). Termination of D-serine signaling is attained by reuptake into astrocytes where it is catabolized by D-amino acid oxidase (DAAO). A1 reactive astrocytes express SR, and synthesize and release D-serine, which may contribute to their neurotoxic effects by acting at extrasynaptic NMDAR. Asc-1, alanine-serine-cysteine-1 (Slc7a10; solute carrier family 7 member 10); ASCT-1, alanine/serine/cysteine/threonine transporter 1 (Slc14; solute carrier Family 1 member 4); GLDC, glycine decarboxylase; GlyT-1, glycine transporter 1; PHGDH, phosphoglycerate 3-dehydrogenase; PSD-95, postsynaptic density protein 95; VGLUT, vesicular glutamate transporter. Adapted from Coyle and Balu²⁸ and Wolosker *et al.*⁴⁹ with permission.



history of drug use. A particularly dramatic instance occurred in Washington, DC, during the fall of 1973. The admission rate for what appeared to be unusually long, severe, and treatment-resistant initial schizophrenic psychoses suddenly tripled in a community mental health center. These patients had all smoked PCP before becoming psychotic, and their presenting picture was at first indistinguishable from a florid schizophrenic episode.^{61,62} With regard to psychotic symptoms, lysergic acid diethylamide (known as 'LSD') generally produces a psychosis in healthy volunteers characterized by distortions in visual perception lasting for only 8–16 h whereas NMDAR antagonists, such as PCP and ketamine, produce not only schizophrenic-like psychosis (including auditory hallucinations), but also negative and cognitive symptoms lasting for periods up to 2 weeks.⁶² Furthermore, LSD administration induces no more severe effects in patients with schizophrenia than in healthy volunteers. By contrast, administration of glutamate antagonists to stabilize patients with schizophrenia exacerbates their symptoms.^{63,64}

Anti-NMDAR encephalitis is now well established as an autoimmune disorder having schizophrenia-like symptoms caused by autoantibodies against an extracellular domain of NMDAR.⁶⁵ The antibodies are considered to lead the internalization of postsynaptic NMDAR clusters to causing NMDAR downregulation. The disorder most commonly presents with abnormal behavior, such as agitation and aggression, abnormal speech, catatonia, seizures, psychosis (including delusions and hallucinations), and cognitive impairments.⁶⁶ It was reported that 6.5% of patients at first presentation of psychosis fulfilled the diagnostic criteria for schizophrenia and were shown to be NMDAR-antibody positive.⁶⁷

Genetic findings

Twin and family studies over the last half-century have provided compelling evidence of the high heritability of schizophrenia, approaching a ratio of 0.8.² However, the pattern of familial risk declined strikingly from identical to fraternal twins, consistent with complex genetics wherein multiple risk genes of modest effect interacting with environmental risk factors cause the phenotype. Early in the 21st century, many studies were carried out to determine if candidate genes, typically linked to the dopamine hypothesis, were significantly associated with the risk for schizophrenia. A number of positive, significant associations were reported, often reinforced by meta-analyses. Unfortunately, experience with studies of common disorders involving

complex genetics indicated that these studies were statistically underpowered, leading to false positive results, further complicated by publication bias (non-publication of negatives results).

Sufficiently powered studies to agnostically search the genome for linkage between sites and risk for schizophrenia requires thousands of subjects to satisfy the high bar of a significance threshold of 5×10^{-8} required for multiple comparisons. Obviously, such large cohorts of patients cannot be acquired at a single clinic but rather require extensive collaborations and sharing of data among many sites throughout the world. A recent genome-wide association study (GWAS) was carried out involving over 100 000 controls and nearly 40 000 subjects with schizophrenia, which identified 108 sites on the genome that met the strict statistical threshold of 5×10^{-8} . Although one site was linked to the dopamine receptor (*DRD2*), several were associated with glutamatergic neurotransmission or downstream mediators: for example, the mGlu₃ (*GRM3*), the GluN2A (*GRIN2A*), SR (*SRR*), and AMPA receptor 1 (*GRI1*).^{68–70}

Furthermore, other studies have revealed that an increased burden of rare gene variants, taking the form of both large copy-number variants (CNVs) and single-nucleotide variants, which often occur as *de novo* mutations, exert significantly larger effects than common single-nucleotide polymorphisms.^{71–74} According to large-scale CNV studies, *de novo* CNV encode the NMDAR and proteins associated with the postsynaptic density with increased risk of schizophrenia.^{75–79} Especially, 11 rare CNV (deletions at 1q21.1, *NRXN1*, 3q29, 15q11.2, 15q13.3, and 22q11.2, and duplications at 1q21.1, 7q11.23, 15q11.2-q13.1, 16p13.1, and 16p11.2) showed highly significant evidence (odds ratio: 2–60) for association with schizophrenia. About 2.5% of patients and 0.9% of controls carry a CNV at one of these loci.⁸⁰ Moreover, nearly all of them are associated with a range of other neurodevelopmental disorders, such as autism spectrum disorder and intellectual disabilities, and some of them are also associated with particular physical disease phenotypes, such as epilepsy, congenital heart disease, microcephaly, and obesity.

Recently, there has been accumulating evidence also supporting a role for rare variants in schizophrenia causation, especially rare loss of function (LoF; nonsense, splice site or frame shift) variants.⁸¹ For example, in the recent mutation screening study for the exonic regions of the NMDAR subunits in schizophrenia and autism spectrum disorder, 40 rare variants were identified and two of them in *GRIN2C* and *GRIN2D* in schizophrenia were rare LoF mutations.⁸¹ These findings support the view that ultra-rare variants with LoF in

glutamatergic pathways, especially NMDAR-related genes, increase the risk of schizophrenia.

Neurophysiological findings

The development of neuroimaging technology has provided a way forward for non-invasive visualization of the brain structure and activity *in vivo*.⁸² Especially, proton magnetic resonance spectroscopy (¹H-MRS), positron-emission tomography (PET), and single-photon emission computed tomography (SPECT) afford researchers the ability to measure brain parameters, such as dopamine and glutamate function in the brain, in living human subjects.

¹H-MRS

¹H-MRS studies have shown that acute administration of ketamine to healthy subjects leads to a significant increase in anterior cingulate glutamine, a putative marker of glutamatergic neurotransmission, and this increase was marginally related to cognitive function.^{83,84} On the other hand, in schizophrenia, a recent meta-analytic study reported that there was a significant elevation in glutamatergic transmission in the limbic system, in glutamate indices in the basal ganglia, glutamine levels in the thalamus, and Glx (glutamate + glutamine) levels in the basal ganglia and medial temporal lobe.⁸⁵ Poels also showed that there was a positive correlation between elevated Glx in the hippocampus and worse executive functioning, global clinical state, and decreased hippocampal volume in unmedicated patients.^{86,87}

Some ¹H-MRS studies have investigated glutathione (γ -L-glutamyl-L-cysteinyl-glycine) levels in the brains of patients with schizophrenia. Glutathione is a tripeptide synthesized from glutamate, cysteine, and glycine. It has an important role in protecting the brain against oxidative stress as an intracellular antioxidant. Furthermore, it modulates the redox site on the NMDAR. Thus, increasing extracellular glutathione levels promotes glutamate-induced depolarization via NMDAR activation. Some studies indicated that glutathione levels in the medial prefrontal cortex (PFC) were lower in schizophrenia than in healthy controls.⁸⁸ On the other hand, higher levels were found in the medial temporal lobe.⁸⁹ However, significant differences were not observed between schizophrenia and healthy subjects in the other studies.^{90,91} Thus, the results are mixed.

One main limitation of glutamate ¹H-MRS studies is that ¹H-MRS provides a total tissue measure and does not distinguish between intracellular or extracellular compartments, or between intra- or extra-neuronal compartments, further limiting the specificity of such measurements. PET and SPECT allow for a more selective measurement of brain neurochemistry than does ¹H-MRS.⁸⁷

PET/SPECT

One ketamine administration study, which used PET, supports the notion of hypersensitivity to NMDAR antagonists in schizophrenia. In this study, ketamine administration caused regional cerebral blood flow elevations reflecting hypermetabolism in frontal and cingulate regions in both the healthy control group and the schizophrenia group. However, in comparisons of the two groups, the schizophrenia group had greater relative blood flow increases in the anterior cingulate, a region suspected of abnormal glutamatergic function in many ¹H-MRS studies.⁹² On the other hand, PET/SPECT ligands for the glutamate binding site, the GMS, or the redox site on the NMDAR are being developed and are still in the subclinical stage. Regarding the PET ligands, [¹¹C] ABP688 and [¹⁸F] FPEB, which bind to an allosteric site on mGlu₅, which co-localizes with NMDAR functionally and has physically close interactions with NMDAR activation, have demonstrated potential as a marker for glutamatergic transmission.⁸⁷ The translocation protein 18 kDa (TSPO) expression, which is elevated with microglial activation, is starting to be used as another target for PET studies in psychiatric disorders suspected to involve microglial activation, including schizophrenia.⁹³ It is known that activated microglia induce reactive astrocytes by cytokines,^{55,94} which

could contribute to glutamatergic dysfunction in schizophrenia via the activation of extrasynaptic NMDAR by releasing D-serine.

Mismatch negativity/P300/gamma band oscillations

Electroencephalography (EEG) is one of the least invasive and easy-to-use methods to monitor neurophysiological brain function with high temporal resolution in people with psychiatric disorders. Magnetoencephalography is also a neuroimaging technique to visualize neural activity with higher spatial resolution than EEG. Mismatch negativity (MMN), P300 (or P3), and gamma band oscillations are electrical phenomena that appear to be informative markers for schizophrenia.

MMN is a negative-going wave in the event-related potential component and is evoked by a deviant stimulus, which is occasionally inserted in a repetitive train of standard stimuli. The onset of the auditory MMN response occurs within 50 ms of the deviant stimulus, and peaks after an additional 100–150 ms.⁹⁵ MMN is thought to reflect auditory sensory memory or pre-attentive processing.⁹⁶ P300 is a positive-going wave in the event-related potential and is elicited during oddball paradigm with a latency of roughly 250–300 ms after deviant stimulus. Its (P3a) component is thought to reflect a re-orienting or covert shifting of attention.^{95,97}

Administration of the non-competitive NMDAR antagonist, ketamine, is known to attenuate MMN and P300 amplitude in healthy subjects.^{98,99} Furthermore, in individuals with high risk, first-episode, or chronic schizophrenia, attenuation of the MMN amplitude has been reported.^{100,101} Moreover, some studies have provided more direct evidence, showing the linkage of these attenuations and glutamate hypothesis in schizophrenia. For example, the plasma levels of glutamatergic amino acids were correlated with MMN amplitude attenuation in subjects in the early stages of psychosis.⁹⁶ Another EEG and MRS study suggested that there were positive correlations between P300 amplitude and both the glutamine/glutamate ratio and the glutamine concentration in the anterior cingulate.¹⁰² Therefore, MMN and P300 amplitude attenuation has been hypothesized to reflect NMDAR hypofunction and has attracted much attention as a biomarker for schizophrenia. During a continuous performance test, there is a highly significant reduction of no-go anteriorization, which was also observed in both first-episode and chronic schizophrenia.^{103,104}

Synchronous neuronal oscillations in the 30–100 Hz range, known as ‘gamma oscillations,’ are normally correlated with performance of a variety of cognitive tasks, including the allocation of attention and working memory.¹⁰⁵ The gamma rhythms are generated by fast-spiking parvalbumin-positive GABAergic interneurons, which are regulated by NMDA-dependent excitatory input. The NMDAR has also been directly implicated in the emergence of the rhythms.¹⁰⁵ In other words, spontaneous gamma rhythms reflect the excitatory/inhibitory balance between excitatory neuron and inhibitory interneurons. The NMDAR antagonists induce schizophrenic cognitive behaviors and increase spontaneous gamma rhythms.^{106–109} Converging evidence from many studies suggests that abnormalities in the synchronized gamma oscillatory activity elicited by a variety of sensory stimuli and cognitive tasks may be an important biomarker for glutamatergic dysfunction in schizophrenia.^{110,111}

Post-mortem neurochemical findings

Studies of cortical NMDAR expression in schizophrenia using human post-mortem brain tissue have revealed variable changes in transcript and protein expression depending on the brain region and receptor subunit examined.¹¹² A recent meta-analysis indicated that the GluN1 subunit (mRNA and protein) in the PFC of the post-mortem brains of subjects with schizophrenia was significantly decreased relative to controls. In the same study, they also conducted a qualitative review about the GluN2 (A, B, and D) and GluN3A subunits and demonstrated no consistent statistically significant changes in cortical mRNA expression or protein levels of these subunits in schizophrenia

compared to controls, with decreasing GluN2C mRNA expression in the PFC.¹¹³ Some studies also suggested a decreased expression of GluN1 mRNA selectively in the dentate gyrus of subjects with schizophrenia compared to controls^{114,115} and decreased expressions in GluN1 and GluN2B subunits (mRNA and protein) in the left hippocampus of schizophrenia compared to the right.^{115–118}

In addition to direct quantification of the individual NMDAR subunits, increasing postsynaptic density of NMDAR in the post-mortem dorsolateral PFC (dlPFC) in schizophrenia was also reported despite decreased NMDAR signaling at the post-receptor level.¹¹⁹ Furthermore, in a quantitative autoradiograph study, NMDAR binding in the anterior cingulate cortex was suggested to be increased in schizophrenia compared to controls due to a postsynaptic compensation for impaired glutamatergic neurotransmission.¹²⁰

There are also numerous abnormalities of NMDA GMS modulators described not only in the brain but also in the periphery of subjects with schizophrenia. In fact, reductions of brain SR and D-serine have been reported in schizophrenia.^{121–123} The level of kynurenic acid, an endogenous GMS antagonist, is elevated in the cerebral spinal fluid and post-mortem brain tissue in schizophrenia.^{124–126}

Moreover, in addition to the NMDAR itself, altered expression of several NMDAR-associated post-synaptic density proteins have been reported in the post-mortem brains of subjects with schizophrenia. Increased mRNA but decreased protein expression has been reported for both PSD-93 and PSD-95 in the anterior cingulate cortex.^{112,127} The expression of PSD-95 mRNA was significantly decreased in the dlPFC, in spite of the fact that it was increased in the occipital cortex.^{128,129} Similarly, the expression of NF-L mRNA was significantly increased in the dlPFC, even though the protein was decreased.¹²⁷ Furthermore, the kainate subtype of glutamate receptor was first measured in the post-mortem brains of subjects with schizophrenia by Nishikawa *et al.*, who reported a 25–50% increase in [3H] kainic acid binding in the PFC. The increased kainate receptor may also reflect a reduced activity at certain glutamatergic synapses and impaired cognitive functions in the PFC.^{130,131} Some studies also reported that the levels of N-acetylaspartylglutamate (NAAG) and glutamate, as well as the activity of glutamate carboxypeptidase II (GCP-II) were altered in schizophrenia. NAAG, which is catabolized by the enzyme GCP-II, has dual roles as an endogenous NMDAR antagonist and mGlu3 agonist. The elevation of NAAG levels and the reduction of glutamate levels, as well as reduced GCP-II activity in schizophrenia support the hypothesis that NAAG-mediated signaling contributes to NMDAR hypofunction in schizophrenia.^{28,39,132–134}

Animal models

Pharmacologic, developmental, and genetic animal models have the potential to provide a platform for advancing the mechanistic understanding of alterations that can lead to schizophrenia and the possibility to compensate for limitations of human post-mortem studies. There is an abundance of data from these animal models that support the hypothesis of NMDAR hypofunction contributing to the pathophysiology of schizophrenia.³⁹ In pharmacological models, administration of NMDAR antagonists or kynurenic acid to animals led to neurochemical, morphological, and cognitive and/or behavioral features similar to what is observed in schizophrenia. Developmental models, given potential environmental risk factors of schizophrenia during the perinatal and/or early postnatal period, lead to abnormal-brain-development-related NMDAR activity and show neuronal alterations and cognitive and/or behavioral features similar to what is observed in schizophrenia. Genetic animal models have also provided a wealth of data suggesting that reduced NMDAR activity can lead to changes in the brain and behavior that is similar to what is observed in schizophrenia.³⁹

For example, mice lacking the enzyme SR (SR^{-/-}), which is encoded in the schizophrenia risk gene SRR, identified in the large genome-wide association study⁶⁹ showed >85% reduction of endogenous D-serine in the cortex and hippocampus and cognitive

impairments associated with schizophrenia.¹³⁵ SR^{-/-} mice also demonstrate a phenotype that closely replicates many aspects of schizophrenia, including enlarged lateral ventricles, cortical atrophy, reduced dendritic length, reduced spine density, downregulation of the cortical fast-spiking parvalbumin-positive GABAergic interneurons, reduced brain-derived neurotrophic factor expression, reduced Akt signaling, and reduced microRNA-132 levels.^{28,39,82,135,136} Mice lacking the synaptic protein dysbindin encoded by *DTNBP1*, which is reduced in dlPFC and the hippocampus of schizophrenia,^{137,138} show a schizophrenia-like phenotype, including NMDAR hypofunction, disrupted inhibitory transmission, hyperexcitability in the PFC, as well as cognitive impairments, such as working memory.³⁹ Other genetic models, such as insertion of *G72* mice¹³⁹ and inactivation of *DAO* mice,¹⁴⁰ are also known to display NMDAR hypofunction and neurochemical, morphological, and cognitive and/or behavioral phenotypes, such as schizophrenia.

Treatment based on Glutamate Hypothesis

Pharmacotherapy based on the dopamine D₂ receptor (D2R) blocking activity of all antipsychotics developed since the discovery of chlorpromazine over 50 years ago has universally been the mainstay of treatment for patients with schizophrenia. The introduction of antipsychotics was associated with dramatic reduction in the number of patients with schizophrenia and related psychotic disorders in chronic mental hospitals. By introducing 5-hydroxytryptamine₂ (5-HT₂)-receptor blocking activity into the structure of antipsychotics, a second generation of antipsychotics (SGA) was developed that had markedly reduced propensity for causing extrapyramidal neurologic side-effects. Clinical studies indicate that delays in treatment with antipsychotics are associated with poorer outcomes. Thus, D2R blocking approach has provided substantial benefits to patients with schizophrenia. However, negative and cognitive symptoms are refractory and are associated with persistent disability.¹⁴¹ Furthermore, risks of troublesome side-effects, such as weight-gain, hyperlipidemia, glucose intolerance, type 2 diabetes, and metabolic syndrome, further detract from current treatments, especially with SGA.

Meta-analyses indicate that clozapine, a weak D2R antagonist, is associated with consistently better outcomes than all other antipsychotics and appears to affect negative symptoms.¹⁴² The mechanism whereby clozapine is significantly more effective remains unclear as clozapine interacts with several receptors aside from the D2R, including muscarinic, histamine, and serotonin receptors. However, recent studies indicate that clozapine may enhance NMDAR function through different mechanisms, including blocking glycine uptake, enhancing GMS occupancy, and interacting with the mGlu₅.^{143–146}

The glycine modulatory site

The GMS of the NMDAR is the first glutamatergic strategy pursued as a novel mechanism to treat schizophrenia, especially the negative and cognitive symptoms. This modulatory approach trumped direct NMDAR agonists because it is associated with a reduced risk of excitotoxicity and neuronal death associated with direct NMDAR agonists.¹⁴⁷ There are several strategies by which the availability or concentration of GMS co-agonists and antagonists can be altered as a means to augment NMDAR function.³⁹

Glycine is an α -amino acid, which also has a role as an inhibitory neurotransmitter via binding to strychnine-sensitive glycine receptors. Furthermore, glycine enhances NMDA channel opening via the GMS of the NMDAR. Some randomized controlled trials (RCT) with oral glycine suggested improvement of negative symptoms.^{148–151} D-serine is known to be an endogenous co-agonist at the NMDAR and induces more activation of NMDAR than glycine. Some RCT with D-serine or D-alanine also reported efficacy for the treatment of negative symptoms.^{152–154}

D-cycloserine (DCS) is an anti-tubercular drug that inhibits bacterial cell wall synthesis. Neuropsychiatric symptoms, such as depression, sedation, psychosis, and seizures, are side-effects that may occur

with high-dose DCS treatment. DCS is also a partial agonist at the GMS with about 50% efficacy. DCS is a less efficient ligand for NMDAR function than the endogenous full agonists, such as glycine and D-serine. At high doses, DCS acts as an antagonist by displacing more efficacious endogenous full agonists, but at moderate doses, DCS facilitates glutamatergic neurotransmission via the NMDAR.¹⁵⁵

RCT of DCS added to antipsychotics produced mixed results. In the DCS added on first-generation antipsychotics trial, negative symptoms were significantly improved but positive and cognitive symptoms were not.¹⁵⁶ On the other hand, when DCS was added to clozapine, negative symptoms worsened by contrast to the addition of glycine¹⁵⁷ or D-serine.¹⁵⁸ In the trial of addition of DCS to risperidone, which is an SGA, DCS was associated with reduction in negative symptoms. However, the degree of improvement appeared to be intermediate between improvement of negative symptoms observed with combination of DCS with first-generation antipsychotics and worsening of negative symptoms observed with combination of DCS with clozapine.¹⁵⁹ This pattern of response suggests that, as a partial agonist at some NMDAR, DCS may attenuate clozapine effects via the activation of the GMS of the NMDAR.¹⁶⁰

A large multi-site RCT of glycine and DCS added to antipsychotics suggested that they were ineffective. But it was a 'failed' trial as there were significant differences in the outcomes among sites.¹⁶⁰ Post-hoc analysis indicated that at inpatient sites where compliance was assured, glycine and DCS significantly reduced negative symptoms. Along this line, Iwata *et al.*¹⁶¹ recently published a meta-analysis of the results of placebo-controlled add-on trials of NMDAR-positive modulators and concluded that they had no effects on cognitive deficits in schizophrenia but did not address negative symptoms, which has been more consistently improved by GMS agonists.¹⁶² Furthermore, they neglected certain confounds, such as desensitization with continuous DCS treatment¹⁶³ or negative interactions of DCS with clozapine because of its effects on endogenous GMS occupancy.^{143–146} Finally, they neglected to address how DCS has been shown to augment cognitive remediation in schizophrenia, a strategy that employs 'real-world' learning.¹⁶⁴

Increasing GMS occupancy would permit greater NMDAR activation by glutamate. One strategy that has been explored is to inhibit the uptake of the co-agonist, glycine. Previous studies in the acute hippocampal slice at the glutamatergic synapse on the CA1 pyramidal neuron revealed that the GMS is not saturated with endogenous co-agonists and that inhibiting the glycine transporter (GlyT-1) potentiates NMDAR currents at a synapse where D-serine is the dominant co-agonist.¹⁶⁵ Thus, blocking GlyT-1 appeared to be a plausible way to enhance NMDAR function in schizophrenia.

Sarcosine, an endogenous GlyT-1 inhibitor, is generated in the process of glycine synthesis. Three RCT of sarcosine added to the stable antipsychotic drug regimen lasting 6 weeks demonstrated that sarcosine was associated with greater reductions in Positive and Negative Syndrome Scale (PANSS) total scores than the placebo and the D-serine group in patients with stable chronic schizophrenia, as well as in drug-naïve patients in the acute phase of schizophrenia. Moreover, a meta-analysis reported sarcosine was effective in total psychopathology, negative symptoms, and general psychopathology. However, several drugs that exploited sarcosine as the 'back-bone' for high-affinity GlyT-1 inhibitors produced undesirable side-effects, such as hypoactivity and ataxia, and have therefore prompted the development of non-sarcosine-based GlyT-1 inhibitors. Unfortunately, the noncompetitive GlyT-1 antagonist, bitopertin, failed to reach its endpoints to improve PANSS total score and negative symptoms in Phase II/III and III trials for schizophrenia,^{166–168} even though it significantly reduced negative symptoms in a Phase IIb study in patients with stable, medicated schizophrenia.^{39,169} Moreover, in an RCT to investigate adjunctive treatment with Org25935, a selective inhibitor of GlyT-1, Org25935 did not differ significantly from placebo in reducing negative symptoms or improving cognitive functioning when administered as adjunctive treatment to SGA.¹⁷⁰ Other types of GlyT-1 inhibitors, such as PF-03463275^{171,172} and ASP2535,¹⁷³ have

shown promise in preclinical studies of cognitive remediation for schizophrenia.

DAAO, the primary enzyme responsible for catabolizing D-serine, is sensitive to inhibition by benzoic acid. Inhibition of DAAO would presumably increase the availability of D-serine. In an RCT to investigate the effects of add-on treatment with sodium benzoate, benzoate significantly improved a variety of symptom domains and neurocognition in patients with chronic schizophrenia.¹⁷⁴ Benzoate is also known to induce the expression of brain-derived neurotrophic factor in primary human neurons and astrocytes,¹⁷⁵ which might be another pathway whereby benzoate could reduce schizophrenic symptoms.

Stimulating the redox modulatory site on the NMDAR also enhances NMDAR function. Therefore, the redox modulatory site is considered to be another potential target for treatment of schizophrenia. Glutathione can protect a brain from reactive oxidative stress and harmful xenobiotics as a nucleophilic scavenger and an enzyme-catalyzed antioxidant. It also induces NMDAR activation via binding to the redox modulatory site. Glutathione is synthesized from three amino acids, L-glutamate, glycine, and L-cysteine, which is regulated by extracellular *N*-acetyl-L-cysteine (NAC) concentration. NAC increases extra-cellular L-glutamate levels and intracellular glutathione levels via cystine-glutamate anti-porter. Moreover, NAC promotes activation of the Group II mGlu receptors. As a result of these complementary mechanisms, NAC can enhance NMDAR function and reduce excitotoxicity.

The results of several clinical trials with NAC in schizophrenic subjects stabilized on antipsychotic drugs have been reported. In one of the earliest RCT, NAC added to SGA improved the PANSS scores (Negative, General, and total scores) and the Clinical Global Impression scales (Severity and Improvement scales) in chronic schizophrenia.¹⁷⁶ In other trials, in which NAC was added to risperidone in chronic schizophrenia¹⁷⁷ or was added onto clozapine in clozapine-resistant schizophrenia,¹⁷⁸ NAC was also reported to be efficacious for reducing negative and cognitive symptoms. Furthermore, one RCT reported efficacy not only with negative and cognitive symptoms but also with positive symptoms in chronic schizophrenia.¹⁷⁹ Moreover, a recent RCT demonstrated continuous effects of adjunctive NAC approach in long-term treatment¹⁷⁹ and effects in early psychosis.¹⁸⁰ A recent meta-analysis from 6 RCT showed that adjunctive NAC appears to have efficacy for schizophrenia.¹⁸¹

Another strategy to reduce negative and cognitive symptoms in schizophrenia would be to treat for excitotoxicity-related neuropathology with adjunctive memantine. Memantine is a moderate-affinity noncompetitive NMDAR antagonist, which binds preferentially to the same site as MK-801 and PCP within the NMDAR channel. Like Mg²⁺, memantine shows a strong voltage dependency. Memantine can enter the channel and block current flow only if the channel is open. Thus, it is defined as an 'open-channel blocker' or a 'trapping-channel blocker' of NMDAR.¹⁸² Memantine is thought to block pathologically activated NMDAR when synaptic glutamate concentrations are abnormally high whereas it does not influence the normal functioning of physiologically activated receptors. In other words, memantine blocks the effects of sustained, pathologically elevated levels of glutamate that would lead to neuronal dysfunction.¹⁸³ A meta-analysis (including eight RCT¹⁸³) and a systematic review article (including 10 studies¹⁸² in which memantine was add onto antipsychotic drugs in schizophrenic patients) were recently published. Both studies concluded that memantine selectively improves negative symptoms while cognitive and positive symptoms were not significantly affected. Moreover, the meta-analysis also demonstrated that the most robust effects on negative symptoms were associated with young adult schizophrenic patients.¹⁸³ However, long-term effects and tolerance of the approach have been unclear.

Metabotropic glutamate receptors

The mGlu receptors are classified into three groups (Group I: mGlu₁ and 5; Group II: mGlu₂ and 3; and Group III: mGlu₄, 6, 7 and 8) differentiated by their amino acid sequence, ligand selectivity, and signaling

cascades. Subtypes 2, 3, and 5 have been investigated as potential therapeutic targets for schizophrenia.

Group II mGlu receptors are widely expressed throughout the brain, particularly in those regions implicated in schizophrenia, including the hippocampus, cortex, nucleus accumbens, striatum, and amygdala.^{39,184} These receptors are expressed presynaptically as autoreceptors, activated by astrocytic glutamate release or glutamate overflow from the synapse during excessive glutamate release. mGlu₃ is also found postsynaptically as well as on astrocytes where it mediates neuroprotective effects and participates in astrocytic-neuronal communication.¹⁸⁵ Group II mGlu receptors also have a reciprocal relationship with 5-HT_{2A} receptors. Activation of the 5-HT_{2A} receptor enhances thalamocortical neurotransmission in rodents and this effect is antagonized by activation of Group II mGlu receptors. On the other hand, orthosteric agonists of mGlu_{2/3} functionally antagonize 5-HT_{2A} receptor signaling.¹⁸⁵

The mGlu₅ is primarily enriched postsynaptically in both GABAergic interneurons and pyramidal neurons in the hippocampus, cortex, striatum, caudate nucleus, nucleus accumbens, septum, and olfactory bulb. The receptor has close interaction with the NMDAR activation via intracellular signaling pathways and scaffolding proteins, such as HOMOR, SHANK, and GKAP-PSD95.¹⁸⁵ Therefore, it would be of great interest to study whether positive allosteric modulators (PAM) of mGlu₅ can treat schizophrenia.

One of the mGlu_{2/3} agonists, LY2140023, showed effects for both positive and negative symptoms compared to placebo without prolactin elevation, extrapyramidal symptoms, or weight gain in a Phase II trial.¹⁸⁶ However, in a follow-up multicenter Phase II study, LY2140023 did not significantly separate from placebo due to the large placebo effect.¹⁸⁷ The active comparator in this trial did not differ from placebo, indicating that this was a failed trial. Another mGlu₂ PAM, AZD8529, also failed to show efficacy in a Phase II trial.¹⁸⁸ As for mGlu₅ agonists, a PAM of mGlu₅, VU0409551, has shown robust cognitive and behavioral effects in several animal models of schizophrenia.^{189,190} Another Phase II trial has recently been initiated.¹⁹¹

The failed clinical trials or highly variable outcomes of drugs directed at the NMDAR hypothesis of schizophrenia indicate that our current approach to drug development that relies on categorical diagnoses will subvert meaningful advances. First, the GWAS results indicate that schizophrenia is a disorder of complex genetics involving more than 100 genes, each of which has modest effects. Thus, some patients may be genetically biased much more towards glutamatergic dysfunction than others. Second, sub-grouping to enrich for patients with likely glutamatergic pathology could be accomplished through genotyping (polygenic scores) and assessment of other informative biomarkers, such as cortical glutamate measured by magnetic resonance.¹⁹² Third, additional strategies to reduce variance would be to: focus on clinics that have ‘real’ patients and not volunteers recruited by advertisements, reduce the number of sites with larger numbers of patients, and focus on the early stages of schizophrenia as chronic patients likely have a different pathology.¹⁹³

Conclusion

In this review, we have marshaled the abundant evidence from pharmacologic challenges, post-mortem studies, brain imaging, and genetics supporting the role of dysregulation of glutamatergic neurotransmission in the pathophysiology of schizophrenia. Of course, this pathology must be understood in terms of its disruption in the function of critical circuits, such as downregulation of the corticolimbic PV + GABAergic, disinhibition of pyramidal neurons, and increased release of striatal dopamine.¹⁹⁴ Furthermore, reduced NMDAR function attenuates corticolimbic spine development on pyramidal neurons resulting in an approximately 30% reduction in glutamatergic synapses.^{82,195} Taken together, this cortical pathology accounts for cognitive and negative symptoms of schizophrenia as well as positive symptoms, a downstream consequence of the cortical pathology.

Elucidating the pathologic circuitry in schizophrenia provides novel targets for therapeutic interventions. First, given that the disinhibition of striatal dopaminergic input, which correlates with psychosis, is a consequence of increased glutamatergic output from the cerebral cortex, it is not surprising that D2R blocking antipsychotics have little impact on cortically determined negative and cognitive symptoms. Secondly, pre-clinical studies with a genetic model of schizophrenia indicate that restoring NMDAR function by correcting D-serine deficits or by augmenting NMDAR responsiveness with an mGlu₅ PAM reverses the schizophrenic-like pathology. Although the results have been inconsistent due to a variety of factors, properly executed placebo-controlled clinical trials with NMDAR GMS agonists added to stable doses of antipsychotics suggest improved negative symptoms and enhanced cognitive remediation. Unfortunately, drugs that addressed alternative sites, such as an mGlu_{2/3} agonist to downregulate glutamate release or inhibition of GlyT-1, showed promise in Phase IIb trials only to falter in the Phase III trials. These failures reflected high placebo responses and the reliance on the categorical diagnosis untethered from any biomarkers that could identify likely responders with glutamatergic risk genes or biomarkers.

John F. Nash, Jr., who was awarded the 1994 Nobel Prize in Economics for his path-breaking thesis research on game theory, developed schizophrenia in his late twenties. Years after its onset, he gave an invited lecture to the World Congress of Psychiatry, stating: ‘I would not treat myself as recovered if I could not produce good things in my work. A remission might not be worthwhile in the end.’ Thus, he reminds us that the ultimate goal is not to develop drugs that suppress certain symptoms, like antipsychotics, but rather drugs that correct the fundamental pathology to restore the sense of pleasure and liveliness of thought that are stolen from the individual with schizophrenia. Mounting evidence suggests that understanding the glutamatergic dysregulation in schizophrenia may provide a way forward in developing ‘curative’ treatments.

Acknowledgments

J.T.C. has received research support from National Institutes of Health grants and Y.U. from the Program for Advancing Strategic International Networks to Accelerate the Circulation of Talented Researchers (S2702) of the Japan Society for the Promotion of Science.

Disclosure statement

Y.U. declares no competing interests. J.T.C. holds a patent on the use of D-serine to treat serious psychiatric disorders that is owned by Massachusetts General Hospital but could yield royalties to J.T.C.

Author contributions

Both Y.U. and J.T.C. wrote the manuscript.

References

1. Lesh TA, Niendam TA, Minzenberg MJ, Carter CS. Cognitive control deficits in schizophrenia: Mechanisms and meaning. *Neuropsychopharmacology* 2011; **36**: 316–338.
2. Hilker R, Helenius D, Fagerlund B *et al.* Heritability of schizophrenia and schizophrenia spectrum based on the Nationwide Danish Twin Register. *Biol. Psychiatry* 2018; **83**: 492–498.
3. Brown AS. The environment and susceptibility to schizophrenia. *Prog. Neurobiol.* 2011; **93**: 23–58.
4. Bergsholm P. Is schizophrenia disappearing? The rise and fall of the diagnosis of functional psychoses: An essay. *BMC Psychiatry* 2016; **16**: 387.
5. Bleuler E, Jung C. Komplexe und Krankheitsursachen bei dementia praecox. *Zentralblatt für Nervenheilkunde Und Psychiatrie* 1908; **31**: 220–227.
6. Maatz A, Hoff P, Angst J. Eugen Bleuler’s schizophrenia: A modern perspective. *Dialogues Clin. Neurosci.* 2015; **17**: 43–49.
7. Bleuler E. *Dementia praecox oder Gruppe der Schizophrenien*. Arts-&-Boeve, Nijmegen, 1911.
8. Leucht S, Leucht C, Huhn M *et al.* Sixty years of placebo-controlled antipsychotic drug trials in acute schizophrenia: Systematic review,

- Bayesian meta-analysis, and meta-regression of efficacy predictors. *Am. J. Psychiatry* 2017; **174**: 927–942.
9. Schneider K. *Clinical Psychopathology*. Grune & Stratton, New York, NY, 1959.
 10. Pope HG Jr, Lipinski JF Jr. Diagnosis in schizophrenia and manic-depressive illness: A reassessment of the specificity of ‘schizophrenic’ symptoms in the light of current research. *Arch. Gen. Psychiatry* 1978; **35**: 811–828.
 11. Delay J, Deniker P, Harl JM. Therapeutic use in psychiatry of phenothiazine of central elective action (4560 RP). *Ann. Med. Psychol.* 1952; **110**: 112–117.
 12. Clough PW. Reserpine in the treatment of neuropsychiatric disorders. *Ann. Intern. Med.* 1955; **43**: 632–637.
 13. Kline NS. Use of Rauwolfia serpentina Benth. In neuropsychiatric conditions. *Ann. N. Y. Acad. Sci.* 1954; **59**: 107–132.
 14. Davis JM. Efficacy of tranquilizing and antidepressant drugs. *Arch. Gen. Psychiatry* 1965; **13**: 552–572.
 15. Casey JF, Lasky JJ, Klett CJ, Hollister LE. Treatment of schizophrenic reactions with phenothiazine derivatives. A comparative study of chlorpromazine, trifluorpromazine, mepazine, prochlorperazine, perphenazine, and phenobarbital. *Am. J. Psychiatry* 1960; **117**: 97–105.
 16. Guttmacher MS. Phenothiazine treatment in acute schizophrenia; effectiveness: The National Institute of Mental Health psychopharmacology service center collaborative study group. *Arch. Gen. Psychiatry* 1964; **10**: 246–261.
 17. Gershon S, Hekimian LJ, Floyd A Jr, Hollister LE. Alpha-methyl-tyrosine (AMT) in schizophrenia. *Psychopharmacologia* 1967; **11**: 189–194.
 18. Carlsson A. A half-century of neurotransmitter research: Impact on neurology and psychiatry (Nobel lecture). *Chembiochem* 2001; **2**: 484–493.
 19. Snyder SH, Banerjee SP, Yamamura HI, Greenberg D. Drugs, neurotransmitters, and schizophrenia. *Science* 1974; **184**: 1243–1253.
 20. Janssen PA, Niemegeers CJ, Schellekens KH. Is it possible to predict the clinical effects of neuroleptic drugs (major tranquilizers) from animal data? I. “Neuroleptic activity spectra” for rats. *Arzneimittelforschung* 1965; **15**: 104–117.
 21. Janssen PA, Niemegeers CJ, Schellekens KH. Is it possible to predict the clinical effects of neuroleptic drugs (major tranquilizers) from animal data? *Arzneimittelforschung* 1966; **16**: 339–346.
 22. Janssen PA, Niemegeers CJ, Schellekens KH, Lenaerts FM. Is it possible to predict the clinical effects of neuroleptic drugs (major tranquilizers) from animal data? IV. An improved experimental design for measuring the inhibitory effects of neuroleptic drugs on amphetamine- or apomorphine-induced “Cheroing” and “agitation” in rats. *Arzneimittelforschung* 1967; **17**: 841–854.
 23. Snyder SH. The dopamine hypothesis of schizophrenia: Focus on the dopamine receptor. *Am. J. Psychiatry* 1976; **133**: 197–202.
 24. Yui K, Ikemoto S, Ishiguro T, Goto K. Studies of amphetamine or methamphetamine psychosis in Japan: Relation of methamphetamine psychosis to schizophrenia. *Ann. N. Y. Acad. Sci.* 2000; **914**: 1–12.
 25. Lim J, Lee SA, Lam M *et al.* The relationship between negative symptom subdomains and cognition. *Psychol. Med.* 2016; **46**: 2169–2177.
 26. Johnstone EC, Owens DG, Bydder GM, Colter N, Crow TJ, Frith CD. The spectrum of structural brain changes in schizophrenia: Age of onset as a predictor of cognitive and clinical impairments and their cerebral correlates. *Psychol. Med.* 1989; **19**: 91–103.
 27. Davis KL, Buchsbaum MS, Shihabuddin L *et al.* Ventricular enlargement in poor-outcome schizophrenia. *Biol. Psychiatry* 1998; **43**: 783–793.
 28. Coyle JT, Balu DT. The role of serine racemase in the pathophysiology of brain disorders. *Adv. Pharmacol.* 2018; **82**: 35–56.
 29. Krystal JH, Karper LP, Seibyl JP *et al.* Subanesthetic effects of the non-competitive NMDA antagonist, ketamine, in humans. Psychotomimetic, perceptual, cognitive, and neuroendocrine responses. *Arch. Gen. Psychiatry* 1994; **51**: 199–214.
 30. Javitt DC, Zukin SR. Recent advances in the phencyclidine model of schizophrenia. *Am. J. Psychiatry* 1991; **148**: 1301–1308.
 31. Anis NA, Berry SC, Burton NR, Lodge D. The dissociative anaesthetics, ketamine and phencyclidine, selectively reduce excitation of central mammalian neurones by N-methyl-aspartate. *Br. J. Pharmacol.* 1983; **79**: 565–575.
 32. Javitt DC. Negative schizophrenic symptomatology and the PCP (phencyclidine) model of schizophrenia. *Hillside J. Clin. Psychiatry* 1987; **9**: 12–35.
 33. Moghaddam B, Krystal JH. Capturing the angel in “angel dust”: Twenty years of translational neuroscience studies of NMDA receptor antagonists in animals and humans. *Schizophr. Bull.* 2012; **38**: 942–949.
 34. Coyle JT. The glutamatergic dysfunction hypothesis for schizophrenia. *Harv. Rev. Psychiatry* 1996; **3**: 241–253.
 35. Paoletti P, Bellone C, Zhou Q. NMDA receptor subunit diversity: Impact on receptor properties, synaptic plasticity and disease. *Nat. Rev. Neurosci.* 2013; **14**: 383–400.
 36. Yao Y, Belcher J, Berger AJ, Mayer ML, Lau AY. Conformational analysis of NMDA receptor GluN1, GluN2, and GluN3 ligand-binding domains reveals subtype-specific characteristics. *Structure* 2013; **21**: 1788–1799.
 37. Lu W, Du J, Goehring A, Gouaux E. Cryo-EM structures of the triheteromeric NMDA receptor and its allosteric modulation. *Science* 2017; **355**: eaal3729.
 38. Sullivan JM, Traynelis SF, Chen HS, Escobar W, Heinemann SF, Lipton SA. Identification of two cysteine residues that are required for redox modulation of the NMDA subtype of glutamate receptor. *Neuron* 1994; **13**: 929–936.
 39. Balu DT. The NMDA receptor and schizophrenia: From pathophysiology to treatment. *Adv. Pharmacol.* 2016; **76**: 351–382.
 40. Corrigan JJ, Srinivasan NG. The occurrence of certain D-amino acids in insects. *Biochemistry* 1966; **5**: 1185–1190.
 41. Hashimoto A, Nishikawa T, Hayashi T *et al.* The presence of free D-serine in rat brain. *FEBS Lett.* 1992; **296**: 33–36.
 42. Neims AH, Zieverink WD, Smilack JD. Distribution of D-amino acid oxidase in bovine and human nervous tissues. *J. Neurochem.* 1966; **13**: 163–168.
 43. Mothet JP, Parent AT, Wolosker H *et al.* D-serine is an endogenous ligand for the glycine site of the N-methyl-D-aspartate receptor. *Proc. Natl. Acad. Sci. U. S. A.* 2000; **97**: 4926–4931.
 44. Schell MJ, Molliver ME, Snyder SH. D-serine, an endogenous synaptic modulator: Localization to astrocytes and glutamate-stimulated release. *Proc. Natl. Acad. Sci. U. S. A.* 1995; **92**: 3948–3952.
 45. Hashimoto A, Nishikawa T, Oka T, Takahashi K. Endogenous D-serine in rat brain: N-methyl-D-aspartate receptor-related distribution and aging. *J. Neurochem.* 1993; **60**: 783–786.
 46. Li Y, Sacchi S, Pollegioni L, Basu AC, Coyle JT, Bolshakov VY. Identity of endogenous NMDAR glycine site agonist in amygdala is determined by synaptic activity level. *Nat. Commun.* 2013; **4**: 1760.
 47. Meunier CN, Dallerac G, Le Roux N *et al.* D-serine and glycine differentially control neurotransmission during visual cortex critical period. *PLoS One* 2016; **11**: e0151233.
 48. Wolosker H, Blackshaw S, Snyder SH. Serine racemase: A glial enzyme synthesizing D-serine to regulate glutamate-N-methyl-D-aspartate neurotransmission. *Proc. Natl. Acad. Sci. U. S. A.* 1999; **96**: 13409–13414.
 49. Wolosker H, Balu DT, Coyle JT. The rise and fall of the d-serine-mediated gliotransmission hypothesis. *Trends Neurosci.* 2016; **39**: 712–721.
 50. Papouin T, Henneberger C, Rusakov DA, Oliet SHR. Astroglial versus neuronal D-serine: Fact checking. *Trends Neurosci.* 2017; **40**: 517–520.
 51. Miya K, Inoue R, Takata Y *et al.* Serine racemase is predominantly localized in neurons in mouse brain. *J. Comp. Neurol.* 2008; **510**: 641–654.
 52. Benneyworth MA, Li Y, Basu AC, Bolshakov VY, Coyle JT. Cell selective conditional null mutations of serine racemase demonstrate a predominate localization in cortical glutamatergic neurons. *Cell. Mol. Neurobiol.* 2012; **32**: 613–624.
 53. Balu DT, Takagi S, Puhl MD, Benneyworth MA, Coyle JT. D-serine and serine racemase are localized to neurons in the adult mouse and human forebrain. *Cell. Mol. Neurobiol.* 2014; **34**: 419–435.
 54. Perez EJ, Tapanes SA, Loris ZB *et al.* Enhanced astrocytic d-serine underlies synaptic damage after traumatic brain injury. *J. Clin. Invest.* 2017; **127**: 3114–3125.
 55. Liddelov SA, Barres BA. Reactive astrocytes: Production, function, and therapeutic potential. *Immunity* 2017; **46**: 957–967.
 56. Li S, Uno Y, Rudolph U *et al.* Astrocytes in primary cultures express serine racemase, synthesize d-serine and acquire A1 reactive astrocyte features. *Biochem. Pharmacol.* 2018; **151**: 245–251.
 57. Yamasaki M, Yamada K, Furuya S, Mitoma J, Hirabayashi Y, Watanabe M. 3-Phosphoglycerate dehydrogenase, a key enzyme for l-serine biosynthesis, is preferentially expressed in the radial glia/astrocyte lineage and olfactory ensheathing glia in the mouse brain. *J. Neurosci.* 2001; **21**: 7691–7704.
 58. Yang JH, Wada A, Yoshida K *et al.* Brain-specific Phgdh deletion reveals a pivotal role for L-serine biosynthesis in controlling the level

- of D-serine, an N-methyl-D-aspartate receptor co-agonist, in adult brain. *J. Biol. Chem.* 2010; **285**: 41380–41390.
59. Ehmsen JT, Ma TM, Sason H *et al.* D-serine in glia and neurons derives from 3-phosphoglycerate dehydrogenase. *J. Neurosci.* 2013; **33**: 12464–12469.
 60. Wolosker H, Radziszewsky I. The serine shuttle between glia and neurons: Implications for neurotransmission and neurodegeneration. *Biochem. Soc. Trans.* 2013; **41**: 1546–1550.
 61. Luisada PV. The phencyclidine psychosis: Phenomenology and treatment. *NIDA Res. Monogr.* 1978; **21**: 241–253.
 62. Snyder SH. Phencyclidine. *Nature* 1980; **285**: 355–356.
 63. Lahti AC, Holcomb HH, Medoff DR, Tamminga CA. Ketamine activates psychosis and alters limbic blood flow in schizophrenia. *Neuroreport* 1995; **6**: 869–872.
 64. Lahti AC, Koffel B, LaPorte D, Tamminga CA. Subanesthetic doses of ketamine stimulate psychosis in schizophrenia. *Neuropsychopharmacology* 1995; **13**: 9–19.
 65. Dalmau J, Tuzun E, Wu HY *et al.* Paraneoplastic anti-N-methyl-D-aspartate receptor encephalitis associated with ovarian teratoma. *Ann. Neurol.* 2007; **61**: 25–36.
 66. Warren N, Siskind D, O’Gorman C. Refining the psychiatric syndrome of anti-N-methyl-d-aspartate receptor encephalitis. *Acta Psychiatr. Scand.* 2018; **138**: 401–408.
 67. Zandi MS, Irani SR, Lang B *et al.* Disease-relevant autoantibodies in first episode schizophrenia. *J. Neurol.* 2011; **258**: 686–688.
 68. Sullivan PF, Daly MJ, O’Donovan M. Genetic architectures of psychiatric disorders: The emerging picture and its implications. *Nat. Rev. Genet.* 2012; **13**: 537–551.
 69. Schizophrenia Working Group of the Psychiatric Genomics C. Biological insights from 108 schizophrenia-associated genetic loci. *Nature* 2014; **511**: 421–427.
 70. Horwitz T, Lam K, Chen Y, Xia Y, Liu C. A decade in psychiatric GWAS research. *Mol. Psychiatry* 2019; **24**: 378–389.
 71. Kimura H, Fujita Y, Kawabata T *et al.* A novel rare variant R292H in RTN4R affects growth cone formation and possibly contributes to schizophrenia susceptibility. *Transl. Psychiatry* 2017; **7**: e1214.
 72. Kushima I, Aleksic B, Nakatochi M *et al.* High-resolution copy number variation analysis of schizophrenia in Japan. *Mol. Psychiatry* 2017; **22**: 430–440.
 73. Pocklington AJ, Rees E, Walters JT *et al.* Novel findings from CNVs implicate inhibitory and excitatory signaling complexes in schizophrenia. *Neuron* 2015; **86**: 1203–1214.
 74. Xu B, Roos JL, Levy S, van Rensburg EJ, Gogos JA, Karayiorgou M. Strong association of de novo copy number mutations with sporadic schizophrenia. *Nat. Genet.* 2008; **40**: 880–885.
 75. Kirov G, Pocklington AJ, Holmans P *et al.* De novo CNV analysis implicates specific abnormalities of postsynaptic signalling complexes in the pathogenesis of schizophrenia. *Mol. Psychiatry* 2012; **17**: 142–153.
 76. Fromer M, Pocklington AJ, Kavanagh DH *et al.* De novo mutations in schizophrenia implicate synaptic networks. *Nature* 2014; **506**: 179–184.
 77. Purcell SM, Moran JL, Fromer M *et al.* A polygenic burden of rare disruptive mutations in schizophrenia. *Nature* 2014; **506**: 185–190.
 78. Husi H, Ward MA, Choudhary JS, Blackstock WP, Grant SG. Proteomic analysis of NMDA receptor-adhesion protein signaling complexes. *Nat. Neurosci.* 2000; **3**: 661–669.
 79. Husi H, Grant SG. Isolation of 2000-kDa complexes of N-methyl-D-aspartate receptor and postsynaptic density 95 from mouse brain. *J. Neurochem.* 2001; **77**: 281–291.
 80. Rees E, Walters JT, Georgieva L *et al.* Analysis of copy number variations at 15 schizophrenia-associated loci. *Br. J. Psychiatry* 2014; **204**: 108–114.
 81. Yu Y, Lin Y, Takasaki Y *et al.* Rare loss of function mutations in N-methyl-D-aspartate glutamate receptors and their contributions to schizophrenia susceptibility. *Transl. Psychiatry* 2018; **8**: 12.
 82. Coyle JT, Balu DT, Puhl MD, Konopaske GT. History of the concept of disconnectivity in schizophrenia. *Harv. Rev. Psychiatry* 2016; **24**: 80–86.
 83. Rowland LM, Bustillo JR, Mullins PG *et al.* Effects of ketamine on anterior cingulate glutamate metabolism in healthy humans: A 4-T proton MRS study. *Am. J. Psychiatry* 2005; **162**: 394–396.
 84. Stone JM, Dietrich C, Edden R *et al.* Ketamine effects on brain GABA and glutamate levels with 1H-MRS: Relationship to ketamine-induced psychopathology. *Mol. Psychiatry* 2012; **17**: 664–665.
 85. Merritt K, Egerton A, Kempton MJ, Taylor MJ, McGuire PK. Nature of glutamate alterations in schizophrenia: A meta-analysis of proton magnetic resonance spectroscopy studies. *JAMA Psychiatry* 2016; **73**: 665–674.
 86. Poels EM, Kegeles LS, Kantrowitz JT *et al.* Glutamatergic abnormalities in schizophrenia: A review of proton MRS findings. *Schizophr. Res.* 2014; **152**: 325–332.
 87. Poels EM, Kegeles LS, Kantrowitz JT *et al.* Imaging glutamate in schizophrenia: Review of findings and implications for drug discovery. *Mol. Psychiatry* 2014; **19**: 20–29.
 88. Do KQ, Trabesinger AH, Kirsten-Kruger M *et al.* Schizophrenia: Glutathione deficit in cerebrospinal fluid and prefrontal cortex in vivo. *Eur. J. Neurosci.* 2000; **12**: 3721–3728.
 89. Wood SJ, Berger GE, Wellard RM *et al.* Medial temporal lobe glutathione concentration in first episode psychosis: A 1H-MRS investigation. *Neurobiol. Dis.* 2009; **33**: 354–357.
 90. Matsuzawa D, Obata T, Shirayama Y *et al.* Negative correlation between brain glutathione level and negative symptoms in schizophrenia: A 3T 1H-MRS study. *PLoS One* 2008; **3**: e1944.
 91. Terpstra M, Vaughan TJ, Ugurbil K, Lim KO, Schulz SC, Gruetter R. Validation of glutathione quantitation from STEAM spectra against edited 1H NMR spectroscopy at 4T: Application to schizophrenia. *MAGMA* 2005; **18**: 276–282.
 92. Holcomb HH, Lahti AC, Medoff DR, Cullen T, Tamminga CA. Effects of noncompetitive NMDA receptor blockade on anterior cingulate cerebral blood flow in volunteers with schizophrenia. *Neuropsychopharmacology* 2005; **30**: 2275–2282.
 93. Guilarte TR. TSPO in diverse CNS pathologies and psychiatric disease: A critical review and a way forward. *Pharmacol. Ther.* 2018; **194**: 44–58.
 94. Liddel SA, Guttenplan KA, Clarke LE *et al.* Neurotoxic reactive astrocytes are induced by activated microglia. *Nature* 2017; **541**: 481–487.
 95. Takahashi H, Rissling AJ, Pascual-Marqui R *et al.* Neural substrates of normal and impaired preattentive sensory discrimination in large cohorts of nonpsychiatric subjects and schizophrenia patients as indexed by MMN and P3a change detection responses. *Neuroimage* 2013; **66**: 594–603.
 96. Nagai T, Kirihara K, Tada M *et al.* Reduced mismatch negativity is associated with increased plasma level of glutamate in first-episode psychosis. *Sci. Rep.* 2017; **7**: 2258.
 97. Picton TW. The P300 wave of the human event-related potential. *J. Clin. Neurophysiol.* 1992; **9**: 456–479.
 98. Umbricht D, Schmid L, Koller R, Vollenweider FX, Hell D, Javitt DC. Ketamine-induced deficits in auditory and visual context-dependent processing in healthy volunteers: Implications for models of cognitive deficits in schizophrenia. *Arch. Gen. Psychiatry* 2000; **57**: 1139–1147.
 99. Oranje B, van Berckel BN, Kemner C, van Ree JM, Kahn RS, Verbaten MN. The effects of a sub-anesthetic dose of ketamine on human selective attention. *Neuropsychopharmacology* 2000; **22**: 293–302.
 100. Nagai T, Tada M, Kirihara K *et al.* Auditory mismatch negativity and P3a in response to duration and frequency changes in the early stages of psychosis. *Schizophr. Res.* 2013; **150**: 547–554.
 101. Naatanen R, Todd J, Schall U. Mismatch negativity (MMN) as biomarker predicting psychosis in clinically at-risk individuals. *Biol. Psychol.* 2016; **116**: 36–40.
 102. Hall MH, Jensen JE, Du F *et al.* Frontal P3 event-related potential is related to brain glutamine/glutamate ratio measured in vivo. *Neuroimage* 2015; **111**: 186–191.
 103. Fallgatter AJ, Muller TJ. Electrophysiological signs of reduced prefrontal response control in schizophrenic patients. *Psychiatry Res.* 2001; **107**: 19–28.
 104. Kleinlogel H, Strik W, Begre S. Increased NoGo-antagonism in first-episode schizophrenia patients during continuous performance test. *Clin. Neurophysiol.* 2007; **118**: 2683–2691.
 105. Carlen M, Meletis K, Siegle JH *et al.* A critical role for NMDA receptors in parvalbumin interneurons for gamma rhythm induction and behavior. *Mol. Psychiatry* 2012; **17**: 537–548.
 106. Pinaud D. N-methyl d-aspartate receptor antagonists ketamine and MK-801 induce wake-related aberrant gamma oscillations in the rat neocortex. *Biol. Psychiatry* 2008; **63**: 730–735.
 107. Hakami T, Jones NC, Tolmacheva EA *et al.* NMDA receptor hypofunction leads to generalized and persistent aberrant gamma oscillations independent of hyperlocomotion and the state of consciousness. *PLoS One* 2009; **4**: e6755.

108. Lazarewicz MT, Ehrlichman RS, Maxwell CR, Gandal MJ, Finkel LH, Siegel SJ. Ketamine modulates theta and gamma oscillations. *J. Cogn. Neurosci.* 2010; **22**: 1452–1464.
109. Ma J, Leung LS. Relation between hippocampal gamma waves and behavioral disturbances induced by phencyclidine and methamphetamine. *Behav. Brain Res.* 2000; **111**: 1–11.
110. Uhlhaas PJ, Singer W. Abnormal neural oscillations and synchrony in schizophrenia. *Nat. Rev. Neurosci.* 2010; **11**: 100–113.
111. Hirano Y, Oribe N, Kanba S, Onitsuka T, Nestor PG, Spencer KM. Spontaneous gamma activity in schizophrenia. *JAMA Psychiatry* 2015; **72**: 813–821.
112. Kristiansen LV, Huerta I, Beneyto M, Meador-Woodruff JH. NMDA receptors and schizophrenia. *Curr. Opin. Pharmacol.* 2007; **7**: 48–55.
113. Catts VS, Lai YL, Weickert CS, Weickert TW, Catts SV. A quantitative review of the postmortem evidence for decreased cortical N-methyl-D-aspartate receptor expression levels in schizophrenia: How can we link molecular abnormalities to mismatch negativity deficits? *Biol. Psychol.* 2016; **116**: 57–67.
114. Gao XM, Sakai K, Roberts RC, Conley RR, Dean B, Tamminga CA. Iontropic glutamate receptors and expression of N-methyl-D-aspartate receptor subunits in subregions of human hippocampus: Effects of schizophrenia. *Am. J. Psychiatry* 2000; **157**: 1141–1149.
115. Law AJ, Deakin JF. Asymmetrical reductions of hippocampal NMDAR1 glutamate receptor mRNA in the psychoses. *Neuroreport* 2001; **12**: 2971–2974.
116. Pilowsky LS, Bressan RA, Stone JM *et al.* First in vivo evidence of an NMDA receptor deficit in medication-free schizophrenic patients. *Mol. Psychiatry* 2006; **11**: 118–119.
117. Vrajova M, Stastny F, Horacek J *et al.* Expression of the hippocampal NMDA receptor GluN1 subunit and its splicing isoforms in schizophrenia: Postmortem study. *Neurochem. Res.* 2010; **35**: 994–1002.
118. Geddes AE, Huang XF, Newell KA. GluN2B protein deficits in the left, but not the right, hippocampus in schizophrenia. *BMC Psychiatry* 2014; **14**: 274.
119. Banerjee A, Wang HY, Borgmann-Winter KE *et al.* Src kinase as a mediator of convergent molecular abnormalities leading to NMDAR hypoactivity in schizophrenia. *Mol. Psychiatry* 2015; **20**: 1091–1100.
120. Zavitsanou K, Ward PB, Huang XF. Selective alterations in ionotropic glutamate receptors in the anterior cingulate cortex in schizophrenia. *Neuropsychopharmacology* 2002; **27**: 826–833.
121. Bendikov I, Nadri C, Amar S *et al.* A CSF and postmortem brain study of D-serine metabolic parameters in schizophrenia. *Schizophr. Res.* 2007; **90**: 41–51.
122. Hashimoto K, Engberg G, Shimizu E, Nordin C, Lindstrom LH, Iyo M. Reduced D-serine to total serine ratio in the cerebrospinal fluid of drug naive schizophrenic patients. *Prog. Neuropsychopharmacol. Biol. Psychiatry* 2005; **29**: 767–769.
123. Morita Y, Ujike H, Tanaka Y *et al.* A genetic variant of the serine racemase gene is associated with schizophrenia. *Biol. Psychiatry* 2007; **61**: 1200–1203.
124. Plitman E, Iwata Y, Caravaggio F *et al.* Kynurenic acid in schizophrenia: A systematic review and meta-analysis. *Schizophr. Bull.* 2017; **43**: 764–777.
125. Erhardt S, Blennow K, Nordin C, Skogh E, Lindstrom LH, Engberg G. Kynurenic acid levels are elevated in the cerebrospinal fluid of patients with schizophrenia. *Neurosci. Lett.* 2001; **313**: 96–98.
126. Swarcz R, Rassoulpour A, Wu HQ, Medoff D, Tamminga CA, Roberts RC. Increased cortical kynurenate content in schizophrenia. *Biol. Psychiatry* 2001; **50**: 521–530.
127. Kristiansen LV, Beneyto M, Haroutunian V, Meador-Woodruff JH. Changes in NMDA receptor subunits and interacting PSD proteins in dorsolateral prefrontal and anterior cingulate cortex indicate abnormal regional expression in schizophrenia. *Mol. Psychiatry* 2006; **11**: 737–747.
128. Dracheva S, Marras SA, Elhakem SL, Kramer FR, Davis KL, Haroutunian V. N-methyl-D-aspartic acid receptor expression in the dorsolateral prefrontal cortex of elderly patients with schizophrenia. *Am. J. Psychiatry* 2001; **158**: 1400–1410.
129. Ohnuma T, Kato H, Arai H, Faull RL, McKenna PJ, Emson PC. Gene expression of PSD95 in prefrontal cortex and hippocampus in schizophrenia. *Neuroreport* 2000; **11**: 3133–3137.
130. Nishikawa T, Takashima M, Toru M. Increased [3H]kainic acid binding in the prefrontal cortex in schizophrenia. *Neurosci. Lett.* 1983; **40**: 245–250.
131. Toru M. Biological research on schizophrenia. *Psychiatry Clin. Neurosci.* 1998; **52**: S170–S172.
132. Tsai G, Passani LA, Slusher BS *et al.* Abnormal excitatory neurotransmitter metabolism in schizophrenic brains. *Arch. Gen. Psychiatry* 1995; **52**: 829–836.
133. Ghose S, Chin R, Gallegos A, Roberts R, Coyle J, Tamminga C. Localization of NAAAG-related gene expression deficits to the anterior hippocampus in schizophrenia. *Schizophr. Res.* 2009; **111**: 131–137.
134. Bergeron R, Coyle JT. NAAAG, NMDA receptor and psychosis. *Curr. Med. Chem.* 2012; **19**: 1360–1364.
135. Basu AC, Tsai GE, Ma CL *et al.* Targeted disruption of serine racemase affects glutamatergic neurotransmission and behavior. *Mol. Psychiatry* 2009; **14**: 719–727.
136. DeVito LM, Balu DT, Kanter BR *et al.* Serine racemase deletion disrupts memory for order and alters cortical dendritic morphology. *Genes Brain Behav.* 2011; **10**: 210–222.
137. Talbot K, Eidem WL, Tinsley CL *et al.* Dysbindin-1 is reduced in intrinsic, glutamatergic terminals of the hippocampal formation in schizophrenia. *J. Clin. Invest.* 2004; **113**: 1353–1363.
138. Burdick KE, Lencz T, Funke B *et al.* Genetic variation in DTNBP1 influences general cognitive ability. *Hum. Mol. Genet.* 2006; **15**: 1563–1568.
139. Otte DM, Bilkei-Gorzo A, Filiou MD *et al.* Behavioral changes in G72/G30 transgenic mice. *Eur. Neuropsychopharmacol.* 2009; **19**: 339–348.
140. Labrie V, Duffy S, Wang W, Barger SW, Baker GB, Roder JC. Genetic inactivation of D-amino acid oxidase enhances extinction and reversal learning in mice. *Learn. Mem.* 2009; **16**: 28–37.
141. Fusar-Poli P, Papanastasiou E, Stahl D *et al.* Treatments of negative symptoms in schizophrenia: Meta-analysis of 168 randomized placebo-controlled trials. *Schizophr. Bull.* 2015; **41**: 892–899.
142. Girgis RR, Phillips MR, Li X *et al.* Clozapine v. chlorpromazine in treatment-naïve, first-episode schizophrenia: 9-year outcomes of a randomised clinical trial. *Br. J. Psychiatry* 2011; **199**: 281–288.
143. Javitt DC, Duncan L, Balla A, Sershen H. Inhibition of system A-mediated glycine transport in cortical synaptosomes by therapeutic concentrations of clozapine: Implications for mechanisms of action. *Mol. Psychiatry* 2005; **10**: 275–287.
144. Schwieler L, Linderholm KR, Nilsson-Todd LK, Erhardt S, Engberg G. Clozapine interacts with the glycine site of the NMDA receptor: Electrophysiological studies of dopamine neurons in the rat ventral tegmental area. *Life Sci.* 2008; **83**: 170–175.
145. Gray L, van den Buuse M, Scarr E, Dean B, Hannan AJ. Clozapine reverses schizophrenia-related behaviours in the metabotropic glutamate receptor 5 knockout mouse: Association with N-methyl-D-aspartic acid receptor up-regulation. *Int. J. Neuropsychopharmacol.* 2009; **12**: 45–60.
146. Veerman SR, Schulte PF, Begemann MJ, Engelsbel F, de Haan L. Clozapine augmented with glutamate modulators in refractory schizophrenia: A review and metaanalysis. *Pharmacopsychiatry* 2014; **47**: 185–194.
147. Lawlor BA, Davis KL. Does modulation of glutamatergic function represent a viable therapeutic strategy in Alzheimer's disease? *Biol. Psychiatry* 1992; **31**: 337–350.
148. Javitt DC, Zylberman I, Zukin SR, Heresco-Levy U, Lindenmayer JP. Amelioration of negative symptoms in schizophrenia by glycine. *Am. J. Psychiatry* 1994; **151**: 1234–1236.
149. Heresco-Levy U, Javitt DC, Ermilov M, Mordel C, Horowitz A, Kelly D. Double-blind, placebo-controlled, crossover trial of glycine adjuvant therapy for treatment-resistant schizophrenia. *Br. J. Psychiatry* 1996; **169**: 610–617.
150. Heresco-Levy U, Javitt DC, Ermilov M, Mordel C, Silipo G, Lichtenstein M. Efficacy of high-dose glycine in the treatment of enduring negative symptoms of schizophrenia. *Arch. Gen. Psychiatry* 1999; **56**: 29–36.
151. Javitt DC, Silipo G, Cienfuegos A *et al.* Adjunctive high-dose glycine in the treatment of schizophrenia. *Int. J. Neuropsychopharmacol.* 2001; **4**: 385–391.
152. Tsai G, Yang P, Chung LC, Lange N, Coyle JT. D-serine added to antipsychotics for the treatment of schizophrenia. *Biol. Psychiatry* 1998; **44**: 1081–1089.
153. Heresco-Levy U, Javitt DC, Ebstein R *et al.* D-serine efficacy as add-on pharmacotherapy to risperidone and olanzapine for treatment-refractory schizophrenia. *Biol. Psychiatry* 2005; **57**: 577–585.
154. Tsai GE, Yang P, Chang YC, Chong MY. D-alanine added to antipsychotics for the treatment of schizophrenia. *Biol. Psychiatry* 2006; **59**: 230–234.

155. Hashimoto K, Malchow B, Falkai P, Schmitt A. Glutamate modulators as potential therapeutic drugs in schizophrenia and affective disorders. *Eur. Arch. Psychiatry Clin. Neurosci.* 2013; **263**: 367–377.
156. Goff DC, Tsai G, Levitt J *et al.* A placebo-controlled trial of D-cycloserine added to conventional neuroleptics in patients with schizophrenia. *Arch. Gen. Psychiatry* 1999; **56**: 21–27.
157. Evins AE, Fitzgerald SM, Wine L, Rosselli R, Goff DC. Placebo-controlled trial of glycine added to clozapine in schizophrenia. *Am. J. Psychiatry* 2000; **157**: 826–828.
158. Tsai GE, Yang P, Chung LC, Tsai IC, Tsai CW, Coyle JT. D-serine added to clozapine for the treatment of schizophrenia. *Am. J. Psychiatry* 1999; **156**: 1822–1825.
159. Evins AE, Amico E, Posever TA, Toker R, Goff DC. D-cycloserine added to risperidone in patients with primary negative symptoms of schizophrenia. *Schizophr. Res.* 2002; **56**: 19–23.
160. Goff D. The therapeutic role of D-cycloserine in schizophrenia. *Adv. Pharmacol.* 2016; **76**: 39–66.
161. Iwata Y, Nakajima S, Suzuki T *et al.* Effects of glutamate positive modulators on cognitive deficits in schizophrenia: A systematic review and meta-analysis of double-blind randomized controlled trials. *Mol. Psychiatry* 2015; **20**: 1151–1160.
162. Tsai GE, Lin PY. Strategies to enhance N-methyl-D-aspartate receptor-mediated neurotransmission in schizophrenia, a critical review and meta-analysis. *Curr. Pharm. Des.* 2010; **16**: 522–537.
163. Goff DC, Cather C, Gottlieb JD *et al.* Once-weekly D-cycloserine effects on negative symptoms and cognition in schizophrenia: An exploratory study. *Schizophr. Res.* 2008; **106**: 320–327.
164. Cain CK, McCue M, Bello I *et al.* D-cycloserine augmentation of cognitive remediation in schizophrenia. *Schizophr. Res.* 2014; **153**: 177–183.
165. Bergeron R, Meyer TM, Coyle JT, Greene RW. Modulation of N-methyl-D-aspartate receptor function by glycine transport. *Proc. Natl. Acad. Sci. U. S. A.* 1998; **95**: 15730–15734.
166. Bugarski-Kirola D, Wang A, Abi-Saab D, Blattler T. A phase II/III trial of bitopertin monotherapy compared with placebo in patients with an acute exacerbation of schizophrenia: Results from the CandleLyte study. *Eur. Neuropsychopharmacol.* 2014; **24**: 1024–1036.
167. Hashimoto K. Targeting of NMDA receptors in new treatments for schizophrenia. *Expert Opin. Ther. Targets* 2014; **18**: 1049–1063.
168. Singer P, Dubroqua S, Yee BK. Inhibition of glycine transporter 1: The yellow brick road to new schizophrenia therapy? *Curr. Pharm. Des.* 2015; **21**: 3771–3787.
169. Umbricht D, Alberati D, Martin-Facklam M *et al.* Effect of bitopertin, a glycine reuptake inhibitor, on negative symptoms of schizophrenia: A randomized, double-blind, proof-of-concept study. *JAMA Psychiatry* 2014; **71**: 637–646.
170. Schoemaker JH, Jansen WT, Schipper J, Szegedi A. The selective glycine uptake inhibitor org 25935 as an adjunctive treatment to atypical antipsychotics in predominant persistent negative symptoms of schizophrenia: Results from the GIANT trial. *J. Clin. Psychopharmacol.* 2014; **34**: 190–198.
171. D'Souza DC, Carson RE, Driesen N *et al.* Dose-related target occupancy and effects on circuitry, behavior, and neuroplasticity of the glycine transporter-1 inhibitor PF-03463275 in healthy and schizophrenia subjects. *Biol. Psychiatry* 2018; **84**: 413–421.
172. Roberts BM, Shaffer CL, Seymour PA, Schmidt CJ, Williams GV, Castner SA. Glycine transporter inhibition reverses ketamine-induced working memory deficits. *Neuroreport* 2010; **21**: 390–394.
173. Harada K, Nakato K, Yarimizu J *et al.* A novel glycine transporter-1 (GlyT1) inhibitor, ASP2535 (4-[3-isopropyl-5-(6-phenyl-3-pyridyl)-4H-1,2,4-triazol-4-yl]-2,1,3-benzoxadiazole), improves cognition in animal models of cognitive impairment in schizophrenia and Alzheimer's disease. *Eur. J. Pharmacol.* 2012; **685**: 59–69.
174. Lane HY, Lin CH, Green MF *et al.* Add-on treatment of benzoate for schizophrenia: A randomized, double-blind, placebo-controlled trial of D-amino acid oxidase inhibitor. *JAMA Psychiatry* 2013; **70**: 1267–1275.
175. Jana A, Modi KK, Roy A, Anderson JA, van Breemen RB, Pahan K. Up-regulation of neurotrophic factors by cinnamon and its metabolite sodium benzoate: Therapeutic implications for neurodegenerative disorders. *J. Neuroimmune Pharmacol.* 2013; **8**: 739–755.
176. Berk M, Copolov D, Dean O *et al.* N-acetyl cysteine as a glutathione precursor for schizophrenia: A double-blind, randomized, placebo-controlled trial. *Biol. Psychiatry* 2008; **64**: 361–368.
177. Farokhnia M, Azarkolah A, Adinehfar F *et al.* N-acetylcysteine as an adjunct to risperidone for treatment of negative symptoms in patients with chronic schizophrenia: A randomized, double-blind, placebo-controlled study. *Clin. Neuropharmacol.* 2013; **36**: 185–192.
178. Rossell SL, Francis PS, Galletly C *et al.* N-acetylcysteine (NAC) in schizophrenia resistant to clozapine: A double blind randomised placebo controlled trial targeting negative symptoms. *BMC Psychiatry* 2016; **16**: 320.
179. Sepehrmanesh Z, Heidary M, Akasheh N, Akbari H, Heidary M. Therapeutic effect of adjunctive N-acetyl cysteine (NAC) on symptoms of chronic schizophrenia: A double-blind, randomized clinical trial. *Prog. Neuropsychopharmacol. Biol. Psychiatry* 2018; **82**: 289–296.
180. Conus P, Seidman LJ, Fournier M *et al.* N-acetylcysteine in a double-blind randomized placebo-controlled trial: Toward biomarker-guided treatment in early psychosis. *Schizophr. Bull.* 2017; **44**: 317–327.
181. Zheng W, Zhang QE, Cai DB *et al.* N-acetylcysteine for major mental disorders: a systematic review and meta-analysis of randomized controlled trials. *Acta Psychiatr. Scand.* 2018; **137**: 391–400.
182. Di Iorio G, Baroni G, Lorusso M, Montemitto C, Spano MC, di Giannantonio M. Efficacy of memantine in schizophrenic patients: A systematic review. *J. Amino Acids* 2017; **2017**: 7021071.
183. Kishi T, Matsuda Y, Iwata N. Memantine add-on to antipsychotic treatment for residual negative and cognitive symptoms of schizophrenia: A meta-analysis. *Psychopharmacology (Berl.)* 2017; **234**: 2113–2125.
184. Hovelso N, Sotty F, Montezinho LP, Pinheiro PS, Herrik KF, Mork A. Therapeutic potential of metabotropic glutamate receptor modulators. *Curr. Neuropharmacol.* 2012; **10**: 12–48.
185. Maksymetz J, Moran SP, Conn PJ. Targeting metabotropic glutamate receptors for novel treatments of schizophrenia. *Mol. Brain* 2017; **10**: 15.
186. Patil ST, Zhang L, Martenyi F *et al.* Activation of mGlu2/3 receptors as a new approach to treat schizophrenia: A randomized phase 2 clinical trial. *Nat. Med.* 2007; **13**: 1102–1107.
187. Kinon BJ, Zhang L, Millen BA *et al.* A multicenter, inpatient, phase 2, double-blind, placebo-controlled dose-ranging study of LY2140023 monohydrate in patients with DSM-IV schizophrenia. *J. Clin. Psychopharmacol.* 2011; **31**: 349–355.
188. Litman RE, Smith MA, Doherty JJ *et al.* AZD8529, a positive allosteric modulator at the mGluR2 receptor, does not improve symptoms in schizophrenia: A proof of principle study. *Schizophr. Res.* 2016; **172**: 152–157.
189. Balu DT, Li Y, Takagi S *et al.* An mGlu5-positive allosteric modulator rescues the neuroplasticity deficits in a genetic model of NMDA receptor hypofunction in schizophrenia. *Neuropsychopharmacology* 2016; **41**: 2052–2061.
190. Rook JM, Xiang Z, Lv X *et al.* Biased mGlu5-positive allosteric modulators provide in vivo efficacy without potentiating mGlu5 modulation of NMDAR currents. *Neuron* 2015; **86**: 1029–1040.
191. Sturm S, Delporte ML, Hadi S *et al.* Results and evaluation of a first-in-human study of RG7342, an mGlu5 positive allosteric modulator, utilizing Bayesian adaptive methods. *Br. J. Clin. Pharmacol.* 2018; **84**: 445–455.
192. Kim SY, Kaufman MJ, Cohen BM *et al.* In vivo brain glycine and glutamate concentrations in patients with first-episode psychosis measured by echo time-averaged proton magnetic resonance spectroscopy at 4T. *Biol. Psychiatry* 2018; **83**: 484–491.
193. Krystal JH, Anticevic A. Toward illness phase-specific pharmacotherapy for schizophrenia. *Biol. Psychiatry* 2015; **78**: 738–740.
194. Lisman JE, Coyle JT, Green RW *et al.* Circuit-based framework for understanding neurotransmitter and risk gene interactions in schizophrenia. *Trends Neurosci.* 2008; **31**: 234–242.
195. Balu DT, Li Y, Puhl MD *et al.* Multiple risk pathways for schizophrenia converge in serine racemase knockout mice, a mouse model of NMDA receptor hypofunction. *Proc. Natl. Acad. Sci. U. S. A.* 2013; **110**: E2400–E2409.